SHORT COMMUNICATION

Linear phase imaging using differential interference contrast microscopy

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Summary

We propose an extension to Nomarski differential interference contrast microscopy that enables isotropic linear phase imaging. The method combines phase shifting, two directions of shear and Fourier-space integration using a modified spiral phase transform. We simulated the method using a phantom object with spatially varying amplitude and phase. Simulated results show good agreement between the final phase image and the object phase, and demonstrate resistance to imaging noise.

Introduction

Differential interference contrast (DIC) is a popular method for easily imaging optical path length changes in microscopic specimens (Pluta, 1989). This allows rapid inspection of unstained objects, such as biological tissues in transmission or surface heights in reflection.

Although DIC is known as a phase imaging technique, four major problems persist: (1) standard DIC systems are qualitative in nature, with a non-linear response to optical path length gradients in the specimen; (2) the DIC output intensity is a mix of amplitude and phase gradient contrast; (3) for many applications it is useful to obtain the actual phase of the specimen, whereas DIC gives a directional phase gradient; and (4) it is desirable that any phase reconstruction method be straightforward, non-iterative and yet robust.

Solving these problems requires a method that:
1 has a linear response to specimen phase gradient,
2 isolates the phase gradient from the object amplitude signal,
3 isotropically integrates the phase gradient to obtain the phase, and
4 is robust and non-iterative.

Several approaches to meeting these goals have been proposed in recent years. Phase shifting DIC is a quantitative optical approach to isolating the phase gradient by shifting the DIC prism bias (Hariharan & Roy, 1996; Cogswell et al., 1997; Xu et al., 2001). Although phase shifting DIC relies on a geometrical optics approximation of DIC imaging in order to isolate the phase, Ishiwata et al. (1996) demonstrated an alternative method for isolating the phase based on a partially coherent model. Their method approximates an integral with respect to the bias, by recording four images each with a different bias, then multiplying the image intensity by the sine of the DIC prism bias, and finally adding the images together to isolate the phase.

Shimada et al. (1990) briefly outline a method that at first glance solves the main three problems of linearity, phase isolation and isotropic integration. They demonstrated phase retrieval from a series of DIC images with changing prism bias and shear direction. However, the details are not clearly specified for their phase shifting and phase integration steps. In addition, their method is designed for reflection DIC, and implicitly assumes a constant object amplitude.
Approaches involving iterative computation that also only partially solve the first three problems include line integration and deconvolution (Kam, 1998), variance filtering and directional integration using iterative energy minimization (Feineigle et al., 1996), and rotational diversity (Preza, 2000). The last of these techniques involves taking several rotated DIC images and combining them using iterative deconvolution. Non-iterative yet anisotropic methods include direct deconvolution of DIC images and combining them using iterative deconvolution. The last of these techniques involves taking several rotated images and combining them using iterative deconvolution.

To date no author has outlined a full method that completely addresses all four problems outlined above. In this communication we detail a combined optical and computational extension of DIC that solves these major problems, resulting in a phase image that is linearly proportional to the object phase and that has a laterally isotropic response to specimen phase.

Method

Our method combines four techniques. The first technique is conventional DIC microscopy. For a complex specimen amplitude of the form \( a(x, y) \exp[i\phi(x, y)] \), the 2D DIC image intensity is given by

\[
f_{2\theta}(x, y) = a_{11}^2 + a_{22}^2 + 2a_{12}a_{21}\cos(\phi_{1x} - \phi_{2x} + 2\theta),
\]

where \( a_{1,1}(x + \Delta x, y) \) and \( a_{2,1}(x - \Delta x, y) \) are the amplitudes for two positions in the object separated by a shear \( 2\Delta x \) set by the DIC Wollaston prism. \( \Delta\phi = \phi_{1x} - \phi_{2x} \) is the corresponding phase difference between those two positions, and \( 2\theta \) is the optical DIC bias (Pluta, 1989; Cogswell & Sheppard, 1992). Here we assume geometrical optics and the Born approximation, but we do not assume a constant object amplitude. A schematic of a DIC microscope is shown in Fig. 1.

The second technique is phase shifting DIC (Hariharan & Roy, 1996; Cogswell et al., 1997; Xu et al., 2001). This technique retrieves a linear phase gradient through phase shifting by rotating the bias \( 2\theta \). The bias may be conveniently set by first inserting a quarter wave plate before the analyser. Rotating the analyser then rotates the bias (Hariharan, 1993). We can then obtain the phase gradient in the \( x \) direction using

\[
\Delta\phi_x = \tan^{-1}\left( \frac{f_{2y} - f_{0y}}{f_{0x} - f_{2x}} \right),
\]

where four DIC images \( f \) have been recorded at biases of \( 2\theta = 0, \pi/2, \pi \) and \( 3\pi/2 \). We have now removed both the object amplitude and vignetting from the signal and obtained a linear phase gradient in the \( x \) direction. This step also removes many potential phase-independent system errors, such as weak spots on the camera or non-uniform illumination. But we have so far only imaged the component of the phase gradient that is parallel with the shear direction (van Munster et al., 1998).

The third technique is to repeat the previous two steps with the shear rotated to obtain \( \Delta\phi_y \). The shear direction may be changed by rotating either the specimen or the DIC prisms by \( 90^\circ \). We note that a recently announced variant of DIC called total interference microscopy (Carl Zeiss, Germany) is designed to allow easy rotation of the shear angle. Combinations of DIC with multiple shear directions and phase shifting have been published previously (Hartman et al., 1980; Shimada et al., 1990; Preza et al., 1998; Preza, 2000). However, in those papers a simpler phase shifting technique was applied that assumed a constant object amplitude.

Using the Fourier shift theorem, we can write down the Fourier transforms of our phase gradients

\[
\Delta\phi_x(x, y) \Leftrightarrow 2i\sin(2\pi\Delta x m)f_x(m, n) \tag{3}
\]

\[
\Delta\phi_y(x, y) \Leftrightarrow 2i\sin(2\pi\Delta y n)f_y(m, n), \tag{4}
\]

where \( m, n \) are the spatial frequency co-ordinates, \( i = \sqrt{-1} \), \( \Leftrightarrow \) denotes a two-dimensional (2D) Fourier transform, and capitalization denotes a Fourier transformed function.

This sets the stage for the fourth technique: using Eqs (3) and (4) to obtain the phase \( \phi(x, y) \). We apply a Fourier-space integration approach that is direct, straightforward and reasonably accurate for images that do not contain discontinuities.
such as biological phase images. We begin by combining the $x$ and $y$ phase gradients to form a complex function

$$g(x,y) = \Delta \phi_x + i \Delta \phi_y.$$ (5)

We then perform a 2D Fourier transform on $g(x,y)$ and apply the Fourier shift theorem to give

$$\Phi(m,n) = \begin{cases} 0 & \text{if } [\sin(2\pi \Delta x m), \sin(2\pi \Delta y n)] = [0,0] \\ \frac{G(m,n)}{H(m,n)} & \text{otherwise} \end{cases}$$ (6)

with

$$H(m,n) = 2i[\sin(2\pi \Delta x m) + i \sin(2\pi \Delta y n)].$$ (7)

An inverse Fourier transform of $\Phi(m,n)$ gives the desired phase $\phi(x,y)$. We note that the field of 2D phase unwrapping deals with a similar problem, which can be solved using a range of direct and iterative methods (Ghiglia & Pritt, 1998; Volkov et al., 2002).

For small shear distances $\Delta x$ we can use $\sin x \approx x$ to approximate Eq. (7) with

$$H_d(m,n) = 4\pi \Delta x (m + in).$$ (8)

This is equivalent to approximating the phase gradients $\Delta \phi_x$ and $\Delta \phi_y$ with the partial derivatives $\partial \phi / \partial x$ and $\partial \phi / \partial y$, and then applying the Fourier derivative theorem.

Summarizing the algorithm steps we have:
1. DIC imaging giving $f$.
2. Phase shifting giving $\Delta \phi_x$.
3. Shear rotation giving $\Delta \phi_y$.
4. Fourier phase integration giving the desired phase $\phi$.

**Results**

We have carried out simulations to evaluate the full method. We used a coherent paraxial imaging model, which has been shown to give reasonably accurate predictions for DIC (Preza et al., 1999). However, extending our model for this simulation to include partial coherence and vectorial diffraction should not pose any fundamental difficulties.

The phantom object we simulated is shown in Fig. 2(a,b), with a transmission amplitude varying from 80% to 100% and a phase varying from 0 waves to 0.3 waves. The illuminating beam was monochromatic with wavelength $\lambda = 550$ nm, imaging the sample through a 0.5-NA lens. The shear of the DIC Wollaston prism was set at $2\Delta x = 2\Delta y = 1 \mu m$.

DIC imaging was modelled using fast Fourier transforms (FFTs) with $1024 \times 1024$ pixels including windowing and padding, with the subsequent image being $363 \times 363$ pixels corresponding to a $25 \times 25 \mu m$ region of the object. DIC was simulated using the pupil functions

$$P_x(m,n) = -2i \sin(\theta + k \Delta x m)$$ (9)

$$P_y(m,n) = -2i \sin(\theta + k \Delta y n)$$ (10)

for shear in the $x$ and $y$ directions, respectively, where $k = 2\pi / \lambda$. We added random noise to the intensity of each simulated DIC image, generated using a uniform distribution scaled to fit between 0 and 10% of the intensity range of the image. An example simulated image is shown in Fig. 2(c). Note the image
contains a mixture of amplitude and phase information, with
the amplitude information geometrically distorted owing to
the asymmetrical pupil function in Eq. (9). We used this
DIC imaging model to simulate and compute steps 2 and 3 of
the algorithm. The phase gradient in the x direction \( \Delta \phi_x \) is shown
in Fig. 2(d).

The final step, Eqs (6) and (7), was carried out using
726 \times 726 pixel FFTs, after mirror reflecting the phase gradient image to reduce edge discontinuity effects significantly, as
reflection was implemented by creating a larger image
before applying a Fourier transform to obtain \( G(m,n) \). We also
windowed \( H(m,n) \) to avoid amplifying high-frequency noise in
the image, by setting \( H(m,n) = 0 \) for spatial frequencies outside
the aperture of the simulated imaging system. Steps 1–3 took
54 s to execute on an AMD Athlon 1.4-GHz PC, and performing
step 4 took 4 s.

The final phase image \( \phi \) is shown in Fig. 2(e). This image
shows we have extracted only the phase from the phantom
object, with no visible corruption by either the object amplitu
de or random noise. After normalizing both the object phase
and the retrieved phase, a normalized image of the error
(Fig. 2f) and a line plot (Fig. 3) show good agreement between
the phase image and the phase of the object, with a maximum
error of 17% at the edge of the image. The mean squared error
is \( 1.5 \times 10^{-3} \).

Discussion

The retrieved phase image is qualitatively excellent. However,
certain errors persist, mostly at the top and bottom edges of
the image. The error in those regions is caused by the intersec
tion of the object with the image boundary. These Fourier edge
artefacts might be avoided when acquiring images experimen
tally by placing the spatially varying parts of the object
tirely within the field of view. However, avoiding such object
clipping is not always possible, which is why we have deliber
ately placed parts of our simulated object across the image
boundary. The edge artefacts could also be removed during
processing by using an improved phase integration technique
at the cost of increased complexity and computation (Ghiglia

The results demonstrate that our method has considerable
resistance to imaging noise. The retrieved phase image has
no streaking artefacts, in contrast with the real-space line
integration techniques described by Kam (1998) and Shimada
et al. (1990). By windowing \( H(m,n) \) at the same spatial
frequency cutoff as that imposed by diffraction imaging, we
quenched any high-frequency artefacts introduced by simu-
lated signal noise and the phase retrieval algorithm.

For simplicity in explaining our algorithm, Eqs (1) and (2)
assume geometrical optics. However, our imaging simulation
included diffraction, which will attenuate high spatial fre-
quencies in the phase gradient and thereby introduce addi
tional error in the retrieved phase. Yet despite our simulated
object phase having a broad spatial frequency spectrum, the
Fourier edge artefacts noted above produced larger errors than
the geometrical optics basis of our algorithm. Supplementing
our method with a deconvolution method that accounts

![Fig. 3. A 1D line plot through Fig. 2, vertically downwards from the image centre. Shown are the object amplitude, object phase, DIC image with shear in the x direction and the final phase image from our algorithm. The last three values have been normalized to enable comparison. The horizontal axis is in micrometres and the vertical axis is in normalized units. The retrieved phase has been effectively isolated from the object amplitude and signal noise. The error due to Fourier edge artefacts increases as the plot moves away from the centre of the image.](image-url)
for the effects of diffraction would produce more accurate results, effectively adding the ability to deal with spatially varying amplitudes to the approach by Preza (2000). Such deconvolution methods are generally iterative, thereby increasing the complexity and computation time relative to our non-iterative method.

Careful consideration of sampling is required to maintain high accuracy. We are assuming that the phase gradient $\Delta \phi$ is not too large. Unless $\Delta \phi < \pi$ the DIC phase signal in Eq. (1) will wrap around. An additional limit is imposed by diffraction (Sprague & Thompson, 1972):

$$\Delta \phi / 2\Delta x < k \sin \alpha.$$  \hspace{1cm} (13)

For the system we have simulated, Eq. (12) is a tighter constraint on $\Delta \phi$ than Eq. (13). The size of the diffraction spot provides a tighter limit on $\Delta \phi$ than the shear distance only if the shear distance $2\Delta x$ is less than half the width of the bright-field point spread function (PSF), where the PSF width is defined as $\lambda / \sin \alpha$ (Sprague & Thompson, 1972). Vignetting will also affect the signal for large phase gradients.

An alternative to linear phase imaging is quantitative phase microscopy (Barty et al., 1998). This method obtains the axial intensity derivative using defocus and converts it to separate amplitude and phase images using the transport of intensity equation (TIE). One important difference to our technique is that the TIE image contrast for fine phase details decreases with higher condenser apertures (Barone-Nugent et al., 2002; Sheppard, 2002), whereas DIC imaging gives the best contrast and resolution at the largest condenser apertures.

It is interesting to note that the phase integration method in Eqs (5–7) is related to the Hilbert transform, especially when expressed in the approximate form in Eq. (8). $H_2$ may be rewritten as

$$H_2(m,n) = 4\pi i \Delta x (m^2 + n^2) \exp[\arctan(n/m)].$$  \hspace{1cm} (14)

Applying the spiral phase term $\exp[\arctan(n/m)]$ in Fourier space has been proposed as a 2D version of the Hilbert transform, which is traditionally defined in 1D only (Larkin et al., 2001). It is also known as a complex Riesz transform. This 2D Hilbert transform is isotropic, as compared with the anisotropic 2D half-plane Hilbert transform outlined by Arnison et al. (2000). Although both the modified spiral phase transform in Eq. (14) and the 2D Hilbert transform proposed by Larkin et al. are isotropic, our modified spiral differs by virtue of the amplitude weighting, present in Eq. (14) as the square root term.

In conclusion, we have detailed an extension of DIC that enables isotropic linear phase imaging using phase shifting, two directions of shear and non-iterative Fourier phase integration incorporating a modified spiral phase transform. Simulated results show good agreement between the final phase image and the object phase, for a 2D phantom object with spatially varying amplitude and phase. The method can in principle be used with any DIC imaging system, with potential applications including biological microscopy, 3D visualization, surface profiling, refractive index profiling and X-ray microscopy (David et al., 2002; Kaulich et al., 2002).

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**References**


